

4.3 BATHYMETRIC DISTRIBUTION OF EPIPELON

4.3.1 Introduction

A cursory inspection of Loch Goil at a range of depths (0-40m) suggested that there are differences between the physical environment on the bed of the loch in the shallow sublittoral zone compared with the deeper regions. The sea bed forms a shallow platform at 0-3m (Fig 17) and this is exposed to wave action, surface warming, and optimal light conditions. Beyond this is a slope; here as the water becomes cooler, light is progressively reduced and there is a transition from a coarse sand to finer muddy sediment. In addition, inspection indicated that the abundance of the epipelton varies with depth. Dense patches of epipellic algae discolour the sea bed in the shallows (approx. 0-12m) but are not evident at greater depths.

An assessment of the bathymetric distribution of the epipellic algae in relation to changes in physical and chemical parameters is essential to an understanding of the ecology of Loch Goil. In particular, because the loch is semi-fjordic, the subtidal region comprises most (96%) of the total area (Admiralty chart no. 3746, Edwards and Sharples, 1986). The bathymetry of Loch Goil has been documented (Admiralty chart no. 3746, Edwards and Sharples, 1986; Edwards *et al*, 1986; Mackay and Halcrow, 1976) but there is a paucity of biological data on the loch (Holt and Davies, 1991; CRPB unpublished data, MacDonald, 1927). In particular, there appears to be no information, in the literature, on the benthic micro-algal communities of Loch Goil or any other sea lochs in the Firth of Clyde. Norton and Milburn (1972) documented the depth distribution of marine macroalgae at eleven sites in Argyll from Oban to the Kyles of Bute. They found no macroalgae deeper than 36m and recorded the greatest variety of species in shallow water sheltered from excessive turbulence.

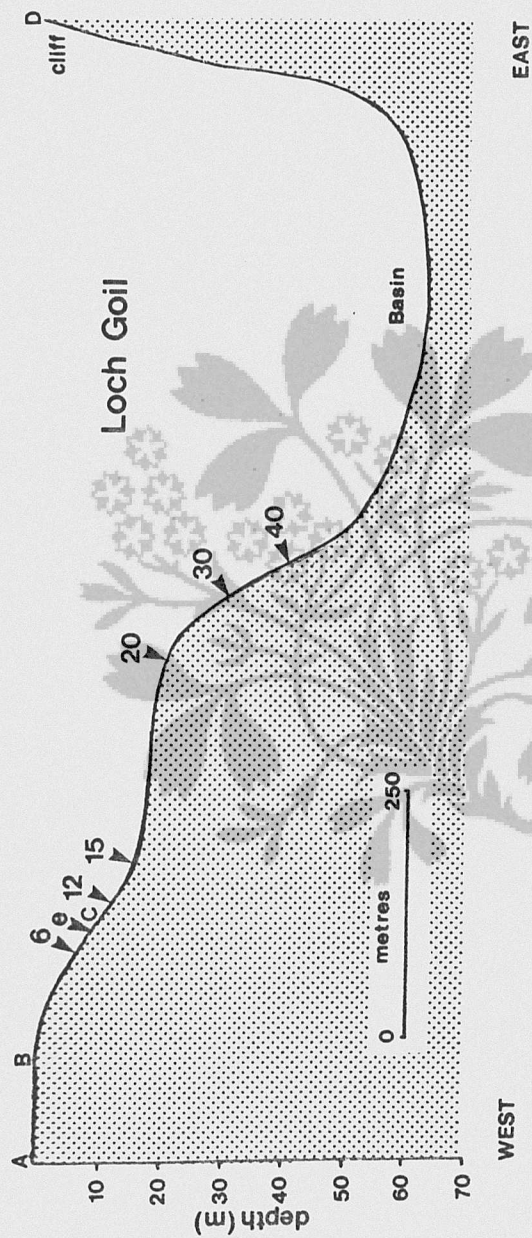


Fig. 17. Cross section of Loch Goil showing the location of the sample stations (6m, 12m, 15m, 20m, 30m, and 40m) along a bathymetric gradient (drawn from Admiralty chart no. 3746). AB is the intertidal zone, A is at the position 56 07.73'N 04 54.45' W; B is the position of Chart Datum; C is the sample site used to monitor seasonal variation of the epipelon; D is at position 56 07.58'N 04 53.38'W.

'e' is the location of the "experimental area" and 9m station used to study the seasonality of the epipelon.

The extent to which the shallow water epipellic community extends into the deep (50-93m) water basin of the Loch Goil is unknown, with little relevant literature from anywhere in Britain or elsewhere. A number of studies have investigated the effect of depth on the epipellic algal community in fresh water lakes (Round, 1961; Roberts and Boylen, 1988, Stevenson *et al*, 1985; Stevenson and Stoermer, 1981; Gruending, 1971; Kingston *et al*, 1983; Stevenson *et al*, 1985) and marine habitats (Bodeanu, 1964, 1968, 1971; Manea and Skolka, 1961; Sundbäck, 1986; Smyth, 1955; Plant-Cuny, 1969, 1973). These studies indicate that bathymetric zonation of species and algal density is a common feature of the sublittoral epipelon although there appears to be no common pattern to the species that inhabit a particular depth or light regime. These changes with depth are attributed, primarily, to variations in the amount and spectral composition of light reaching the sediment. According to Round (1981):

"other factors such as wave sediment, grazing etc., undoubtedly interact with light in determining the distribution (...of species, biomass and photosynthesis of epipelon) under various depths of water." "other factors

The effects of light intensity on benthic micro algae has been investigated extensively in field and laboratory studies, which are reviewed by Admiraal (1984) and Colijn and van Buurt (1975).

Smyth (1955) investigated the depth distribution of epipellic algae in Loch Sween (Argyll), but only as a minor part of a broader ecological investigation of the benthic diatoms of Loch Sween. Smyth used glass slides laid on the surface of the mud from 1-20m depth, at 40m depth and in Loch Fyne at 110m. Between 1-10m there was no evidence that depth affected the composition of the diatom flora and differences recorded from 20m were attributed to:

"unusually unfavourable conditions in the mud....that reduced light intensity"

The major species between 1-20m depth included: *Amphora coffeaeformis*, *A. laevis*, *A. macilenta*, *A. marina*, *Cocconeis scutellum*, *Navicula bahusiensis*, *N. pygmaea* and *Nitzschia closterium*. However, Smyth did record a marked difference between the epipellic diatom community in the shallow subtidal region (1-20m) and the deeper water community (40m). At 40m in Loch Sween and at 110m in Loch Fyne small individuals of *Cocconeis scutellum* were observed. In addition a single specimen of *Navicula pygmaea* was recorded at 40m in Loch Sween. Although Smyth recorded these algae from deep water sediments his samples were only qualitative. He also confirmed that only some of the diatoms had healthy chloroplasts and did not succeed in demonstrating that the algae were alive at these depths. Smyth attempted to confirm his observations from slides, by examining core samples. Cores taken at Millport in April 1948 at depths of 20, 40 and 60 fathoms showed there to be only empty frustules of *Navicula abrupta*. An estimate of epipellic abundance made from two core samples taken in Loch Sween showed there to be 1 and 5×10^8 cell m^{-2} at 40m depth. The presence of *Cocconeis scutellum* in deep water sediment is confirmed by Mare, 1942 who recorded this species in abundance at 72m depth in the English Channel.

Sundbäck (1986) studied the epipelon from 5-20m depth in a shallow inlet of the heavily eutrophic Kattegat. From laboratory experiments and field observations Sundbäck showed that light-saturated photosynthesis occurred down to 10m during April-Sept. and that primary productivity was greatest at 14-16m depth. Benthic microalgae were shown to be limited by both light and nutrients and that microalgal activity influences nutrient fluxes and oxygen conditions at the sediment/water interface.

Plante-Cuny (1973) investigated the bathymetric distribution (5-38m) of diatoms living in tropical marine sands off the NW coast of Madagascar. He found that mean cell sizes of epipellic diatom populations increased as

sediment particle size decreased and as depth increased. The distribution of sediment with depth was linked to a hydrodynamic gradient.

This study of the bathymetric distribution of epipelon in Loch Goil is particularly interesting as it represents the first quantitative investigation of its kind in a sea water loch. The unusual epipellic community of Loch Goil (see Appendix 1) and the unique hydrodynamical characteristics of sea lochs in general (compared with fresh water lakes or coastal and oceanic marine waters) reduces the relevance of the literature in predicting the pattern of depth distribution in this habitat.

Therefore, the objectives of this study were (1) to estimate the abundance and biomass of the epipelon along a gradient of depth, (2) describe the bathymetric distribution of epipellic species, (3) describe the temporal variation of this bathymetric distribution, recorded in the summer of two consecutive years, (4) determine whether epipellic algae inhabit the deep water basin of Loch Goil to a depth of 40m and (5) attempt to relate the changes in the epipelon to light penetration, sediment type, and to the gradient of physical and chemical parameters.

4.3.2 Methods

4.3.2.1 Sampling methods

Replicate sediment samples ($9 \times 15.2 \text{ cm}^2$) were collected by hand at depths of between 6m and 40m in June 1989 and July 1990. Sampling was by a team of four divers operating from an inflatable boat. Fig. 17. shows the location of the sample sites along a transect which extended approx. 500m from the western shore at $56^{\circ} 07.69' \text{N}$, $04^{\circ} 54.33' \text{W}$ (Admiralty chart no. 3746). The relocation of the sample stations was achieved using an echo-sounder and sextant.

Samples of interstitial water were collected at the sediment/water interface using a polypropylene syringe (50ml) and the concentration of dissolved oxygen (DO), salinity and temperature were measured (see 3.4). An attempt to measure the light attenuation with depth was made using a portable monitoring unit (see 3.4.1.2). However, in both 1989 and 1990 this proved unsuccessful due to technical difficulties with the apparatus and insensitivity of the probe to light intensity at depths greater than 6m. However, having observed the levels of light over the whole depth profile I can confirm that no discernible light penetrates to depths greater than approx. 30m depth. Therefore, a conservative estimate would be that 30m to 40m depth represents the lower limit of the euphotic zone within Loch Goil.

4.3.2.2 Laboratory methods

Epipelagic algae were harvested from the samples using the standard extraction method (see 3.2). Living cells (intact chloroplasts) were counted and an estimate made of epipelagic algal abundance, chlorophyll-a and composition of species (see 3.3).

Sediment particle size analysis was carried out on the samples following the extraction of epipelagic algae (see 3.4.6) and the data characterized by calculating the mode, median, mean and sorting statistic (McManus, 1988).

4.3.2.3 Data analyses

Multivariate statistical analyses (Dendrogram, Twinspan and Decorana) were used to compare the epipelagic species abundance data recorded from the different depths, and provide quantitative estimates of dissimilarity between sample communities (see 3.5.1). Spearman Rank correlation analysis was used to analyse relationships between scores from the first two ordination axes (Decorana), epipelagic density and environmental variable values. These analyses were then used as a descriptive tool to discern basic patterns of distribution and to test

the null hypothesis (H_0): "there are no differences in the epipelon along a bathymetric gradient in Loch Goil". The use of multivariate and rank correlation analyses on a large and complex data set, is similar to the analysis methodology used by Blanc *et al* (1972) for the study of marine phytoplankton.

Graphs are used to compare community data recorded in 1989 with that recorded in 1990 and to relate changes in the epipelon in both years to the gradient of physical and chemical parameters.

4.3.2.4 Sampling and analysis of the phytoplankton

A composite sample of phytoplankton was collected from a vertical column (1.13m²) of water to a depth of 8.2m. This was achieved by means of a length of plastic hose (internal diameter 12mm) which was weighted at one end and hung over the side of the stationary boat. The top of the hose was then blocked and the weighted end retrieved by means of a line. The sample (1l) was transferred to a container and preserved with Lugol's Iodine. In the laboratory the phytoplankton was allowed to precipitate in the dark for approx. 12hrs. The precipitate was then mixed and an aliquot (4 drops) removed for qualitative analysis of the phytoplankton community.

4.3.3 Results

4.3.3.1 Epipellic Abundance and Biomass

The epipelon showed a dramatic decrease in abundance and chlorophyll-a (biomass) with increasing depth in both 1989 and 1990 (Fig 18).

In June 1989 the abundance of epipelon was negatively correlated ($P < 0.050$) with depth and decreased in a linear fashion with increasing depth. The maximum standing crop (277.84×10^6 cells m⁻²) was recorded at the shallowest station (6m), decreasing to 1.45×10^6 cells m⁻² at a depth of 30m. No algae (numbers or chlorophyll-a) were found at the deepest station (-40m) by the standard

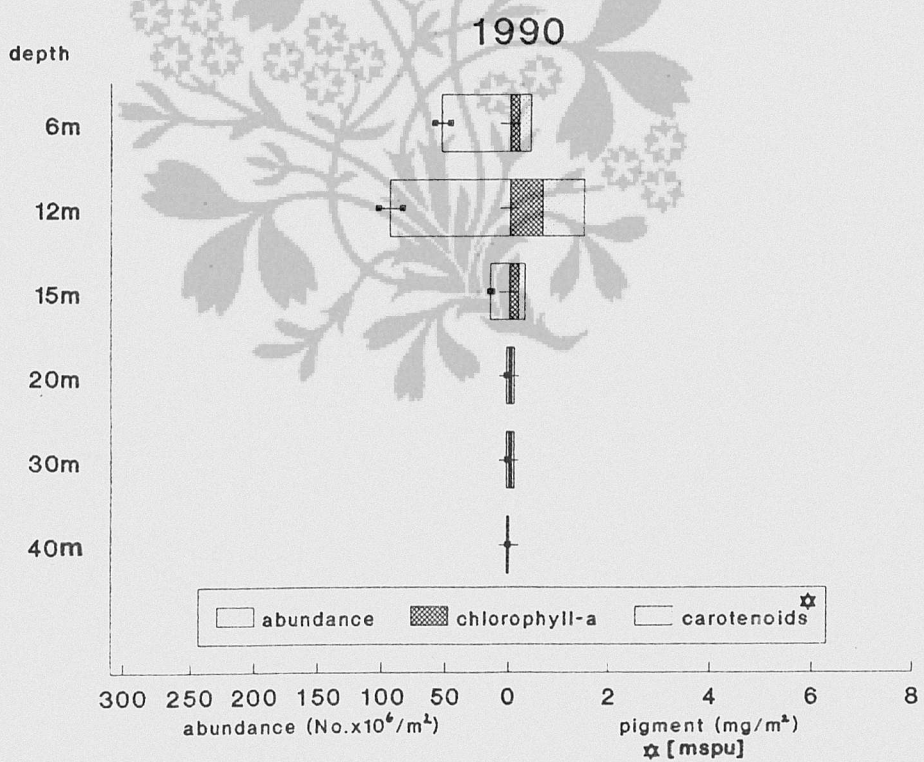
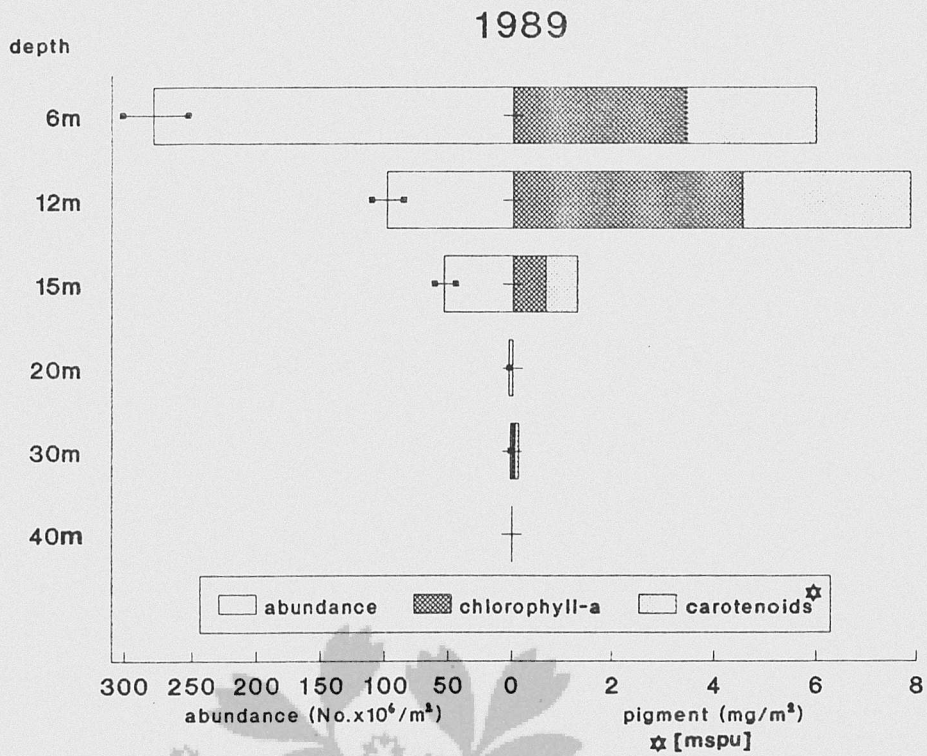


Fig. 18. Distribution of estimated diatom abundance and biomass (pigments) along a depth gradient in June 1989 and July 1990. (Bars indicate standard error).

enumeration procedure. However, 21 live algal cells were extracted from the sample using the standard lens tissue extraction method (see section 3.2.2.4) used to aid taxonomic identification.

The greatest reduction in abundance, in relation to both spatial and vertical separation of the sample stations, was from 6m to 10m. This is consistent with observed reductions in temperature (Fig. 27) and salinity (Fig. 28) and expected attenuation of light intensity. Levels of chlorophyll-a and carotenoids are positively correlated ($P < 0.050$), but their bathymetric distribution indicates a peak in the epipelagic biomass at 12m depth ($4.545 \text{ mg chl-a.m}^{-2}$; $3.296 \text{ MSPU carotenoids m}^{-2}$) whereas the greatest density of algae was recorded at 6m.

In July 1990 there were notably fewer epipelagic algae recorded along the bathymetric profile of Loch Goil compared with the previous year (Fig. 18). Epipelagic abundance was negatively correlated ($P < 0.050$) with depth, however the maximum standing crop ($98.726 \times 10^6 \text{ cells m}^{-2}$) was recorded at a depth of 12m depth, decreasing to $1.11 \times 10^6 \text{ cells m}^{-2}$ at a depth of 40m. Chlorophyll-a, carotenoid and abundance data are all positively correlated ($P < 0.050$) and show a similar bathymetric distribution.

4.3.3.2 Epipelagic Community Analysis

There was generally a very diverse assemblage of epipelagic algae recorded over the two years, comprising 108 taxa representing 34 genera (Tables 6 and 7). The epipelagic algal communities were dominated by diatoms, with one species of silicoflagellate (Chrysophyceae).

4.3.3.2.1 June 1989

Cluster analysis of the 1989 sample community data (Fig. 19) shows the epipelagic communities from the three shallowest sites (6m, 12m, 15m) to be very similar whereas the 20m and 30m communities are clearly grouped as separate. The species composition recorded at 40m depth is



Fig. 19. Dendrograms drawn using group average sorting (Bray-Curtis coefficient) showing the similarity of the epipelagic communities recorded along a bathymetric gradient in Loch Goll, 1989 and 1990.

substantially different from the two main clusters. However, this is attributed to the small number of cells (21 cells) used to estimate the community structure at this station.

The ordination plots (Fig. 20, 1989) from the DECORANA analysis, separates the sample communities into shallow (6m, 12m, 15m) and deep (20m, 30m, 40m) community types along axis 1, the stongest gradient (eigenvalue = 0.291, 82.67 % of the variability). Similarly TWINSpan analysis shows the same separation of sample communities. Examination of the TWINSpan matrix (Fig. 21, 1989) reveals the reason for this grouping, with several of the taxa showing highly restricted distributions along the bathymetric gradient. For example: *Navicula crucigera*, *Nitzschia distans*, *Tropidoneis vitrea* and *Amphora plicata* were only recorded within the shallow subtidal region (6-15m). However, *Nitzschia hybrida* cf. var. *pellucida*, *Tropidoneis lepidoptera*, *Pinnularia divergens* are examples of taxa recorded only at the deeper stations (20-40m).

The bathymetric distribution pattern of the sample communities are seen in Table 6. and the 24 most abundant taxa are shown graphically in Fig. 22. These data show that despite some differences in the community with depth, the most abundant taxa (*Navicula* cf. *tripunctata*, *Navicula forcipata* var. *densistriata*, *Navicula distans*, *Navicula arenaria*, *Navicula* sp.F/G/H, *Amphora* cf. *proteus* var. B) appear to form the greatest proportion of the sample communities along the entire bathymetric gradient (6-40m). This is summarised in Fig.23 (A) which divides the sample communities into the relative proportions of the major genera. This graph also highlights the increase in the proportion within the sample community of *Nitzschia* sp B/C at 20m and 30m depth and the planktonic diatom *Skeletonema costatum* at 30m and 40m depth.

The approximately linear decrease in epipelagic numbers and chlorophyll-a with increasing depth (Fig. 18, 1989) can be attributed to diminishing abundance of all the

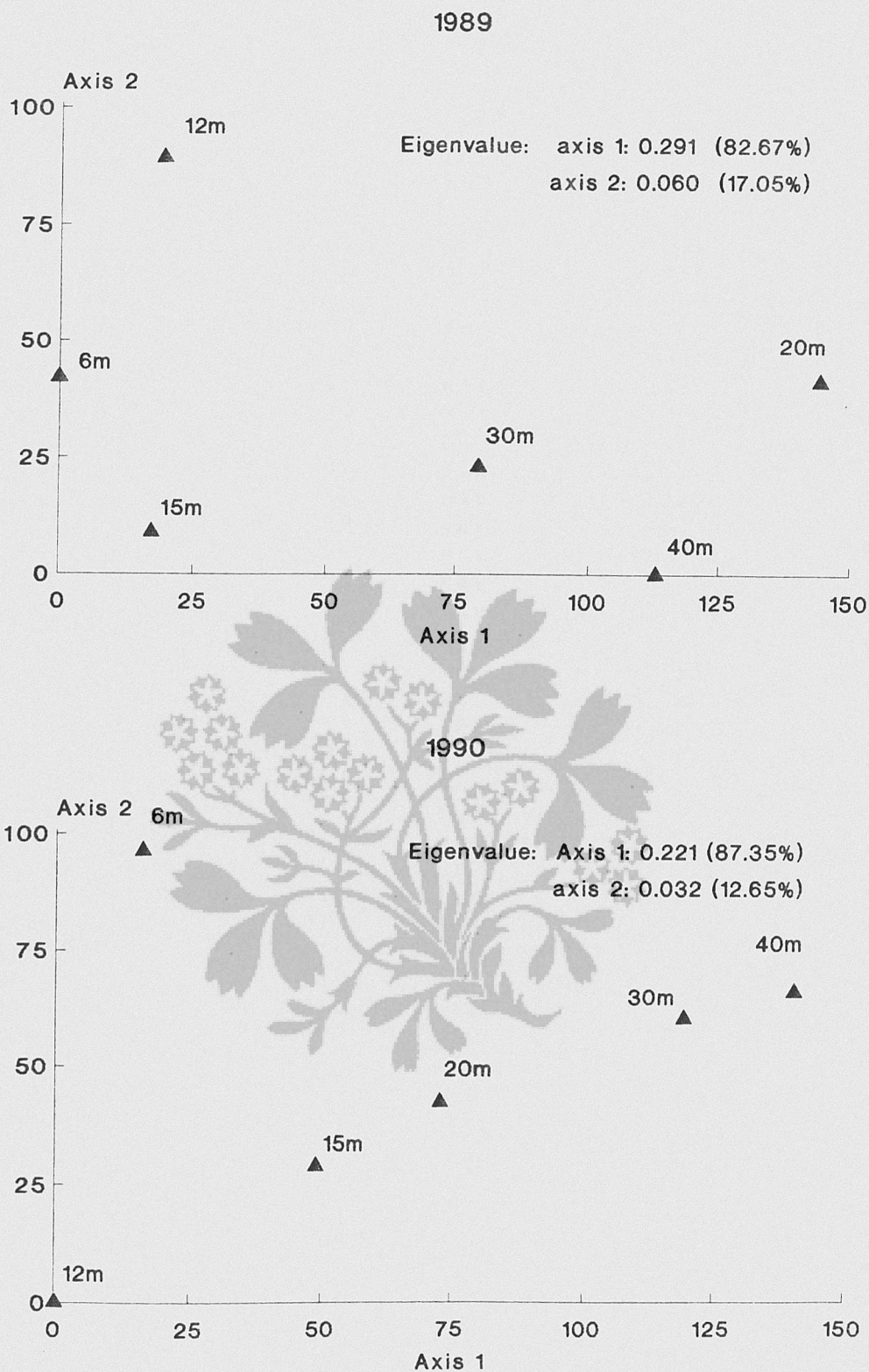
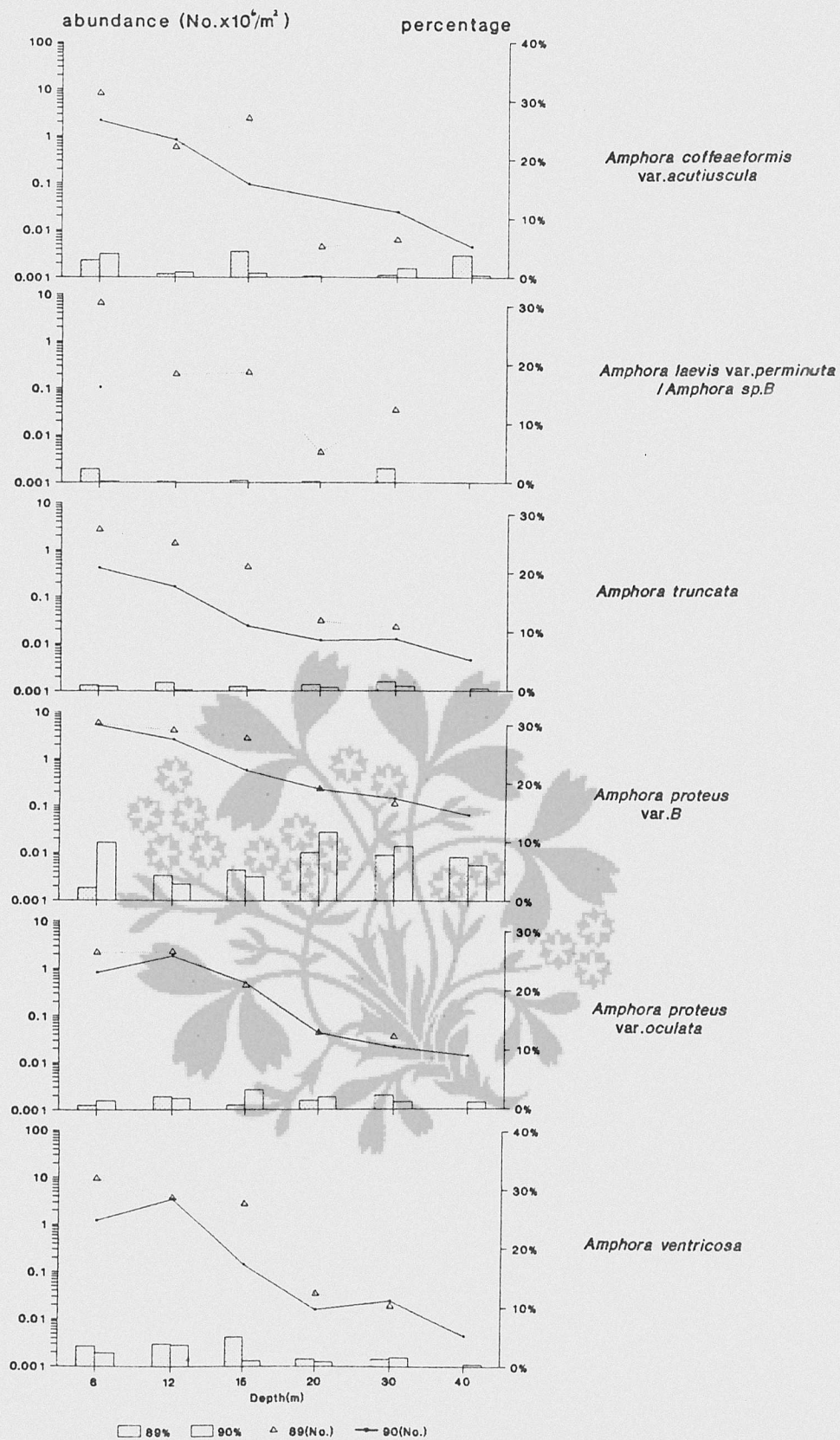


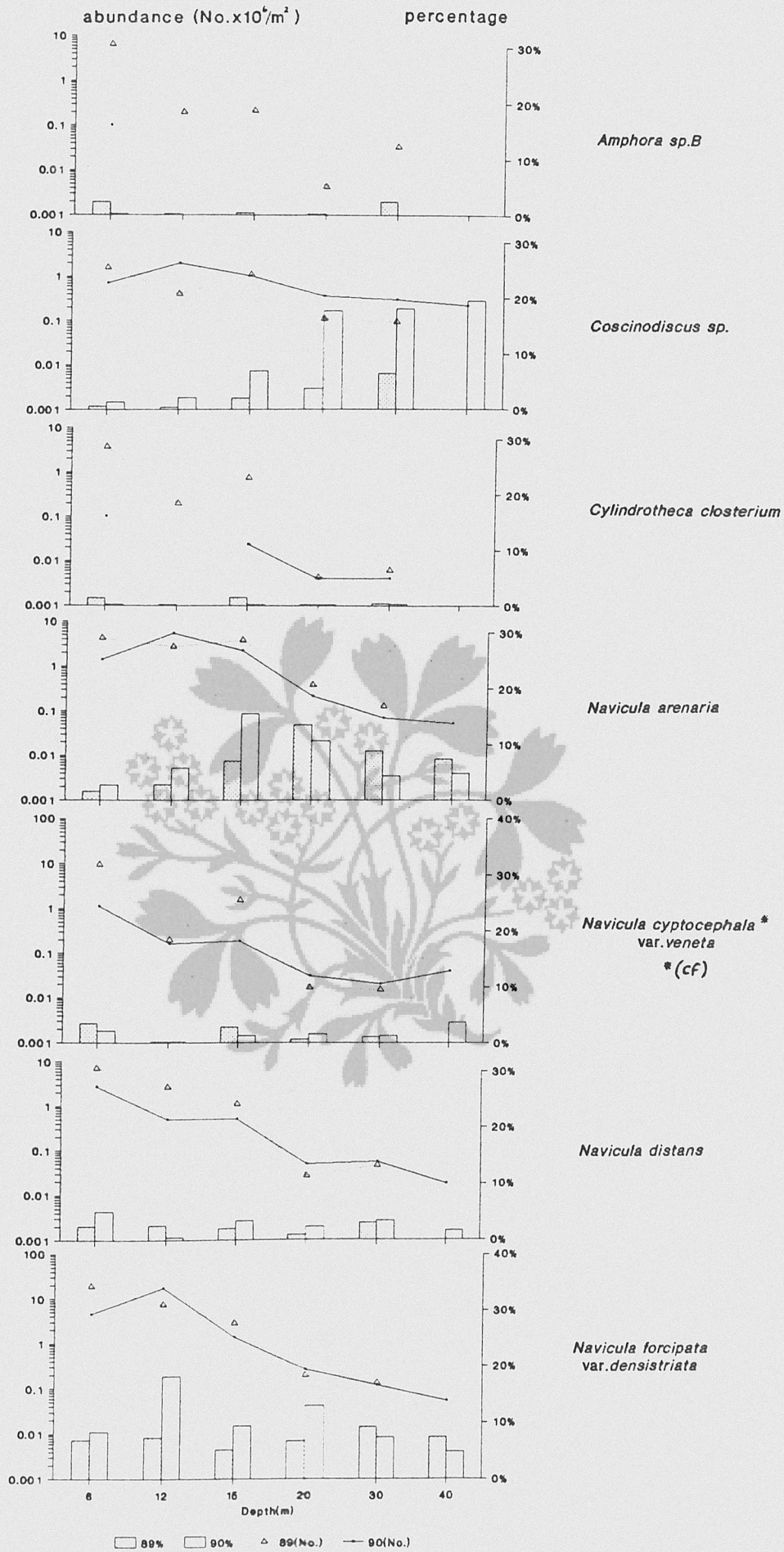
Fig. 20. Ordination plots for the epipelagic community recorded from a series of depths (m) in Loch Goil during 1989 and 1990. Axis 1 and axis 2 data are derived from detrended correspondence analysis (DECORANA). The Eigenvalue and percentage variability are shown for each axis.

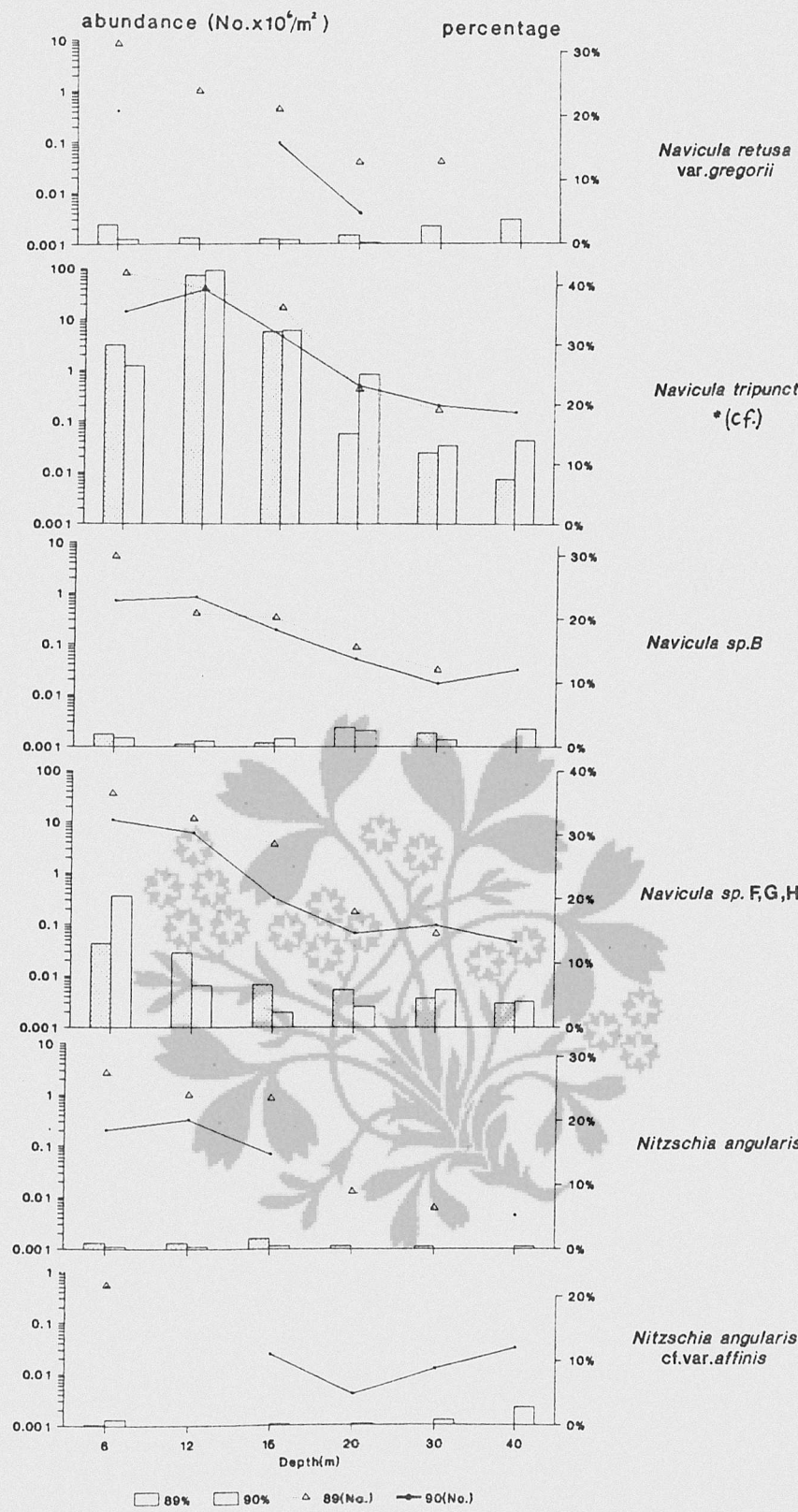
Depth (m)	12 15 6 2030 40				12 15 6 2030 40			
	12	15	6	2030 40	12	15	6	2030 40
STEN	ONSP	-13	---	000	NAVI	GREG	-212	-- 011
NAVI	DICY	-1	----	000	NAVI	DIST	44434	- 011
RHOI	ABBR	1	-----	000	NAVI	CSPF	555551	011
NAVI	CRUC	211	---	000	DIPL	AESF	11111-	011
STEN	INCO	--1	---	000	DIPL	OVAL	21112-	011
ACNA	CONS	--1	---	000	COCC	SCUT	221111	011
EUNO	TISP	--1	---	000	AMPH	OSPB	-2414-	011
CAMP	FAST	11	----	000	AMPH	LAEV	11221-	011
AMPH	OSPC	--1	---	000	AMPH	OSTR	-2212-	011
NITZ	SIGI	-22	---	000	AMPH	PROO	43344-	011
NITZ	DIST	111	---	000	AMPH	TRUN	32333-	011
NITZ	MARG	--2	---	000	AMPH	PROT	554552	011
NITZ	BILO	-1	----	000	NITZ	SSPB	243551	011
NITZ	SIGM	2	-----	000	NITZ	ANGU	333222	011
NITZ	ANGA	--1	---	000	NITZ	PAND	21222-	011
PLEU	NAVI	11	----	000	DONK	RETI	1-211-	011
GYRO	OIST	-11	---	000	STAU	SALI	11112-	10
AMPH	PLIC	2-1	---	000	NAVI	PALP	-1222-	10
TROP	VITR	-22	---	000	NAVI	CSPB	22454-	10
GYRO	TENU	-11	---	000	NAVI	RETG	324341	10
CALO	LIBE	1111	--	001	NAVI	ATLA	-2--2-	10
DIPL	AESD	111-1	-	001	NAVI	HENN	2--11-	10
SURI	FAST	1111	--	001	ACNA	NTHE	--2-2-	10
OPEP	MART	1111	--	001	NAVI	AREN	453552	10
AMPH	OSPA	1-2-1	-	001	ACNA	FIMB	--2-2-	10
AMPH	ARCU	2-11	--	001	TRAC	ASPE	11121-	10
NITZ	INSM	212-1	-	001	SYNE	TABU	--11--	10
PLEU	FORM	1111	--	001	AMPH	PROP	-1-1--	10
PLEU	AUST	21--1	-	001	EUNO	SEPT	-1-1--	10
NAVI	CRYP	14423	-	010	GYRO	ALGO	2-222-	10
NAVI	CSPC	42332	-	010	DICT	SPEC	111111	10
NAVI	BRYO	-2--1	-	010	SKEL	COST	332442	10
DIPL	SMIT	122-2	-	010	ACNA	ANGU	--111-	110
TABE	FLOC	11--1	-	010	COSC	INSP	242553	110
DIPL	SPLE	11--1	-	010	NAVI	PLIC	----1-	111
AMPH	VENT	45433	-	010	PINN	USPA	----1-	111
AMPH	COFF	254121	-	010	PINN	DIVE	----1--	111
NITZ	SOCI	23212	-	010	PINN	RUPE	----1-	111
PLEU	AEST	23322	-	010	SURI	HYBR	----1-	111
PLEU	STRI	22212	-	010	DIPL	CRAB	----1-	111
PLEU	SALI	22-11	-	010	PLAG	STAU	----1--	111
PLAG	TAYR	-111	--	010	FRUS	RHOM	----1-	111
CYLI	CLOS	13312	-	010	NITZ	HYBR	---4-	111
NAVI	CSPA	32323	-	011	TROP	LEPI	---11-	111
NAVI	TRIP	555552	-	011				
NAVI	FDRC	555552	-	011				
NAVI	PHYL	12313	-	011			000111	

Fig. 21.

1989: TWINSPAN classification of epiplic communities recorded in Loch Goil at a range of depths in 1989. Abbreviations of species names are shown at the left; sample numbers along the top. The classification of species and samples are indicated along the right and bottom margins (Hill, 1979 [a]). The main dichotomy for the samples is indicated by a vertical separation. Values indicate a scale of abundance, with absence of a species represented by the symbol "-".







89% 90% △ 89(No.) — 90(No.)

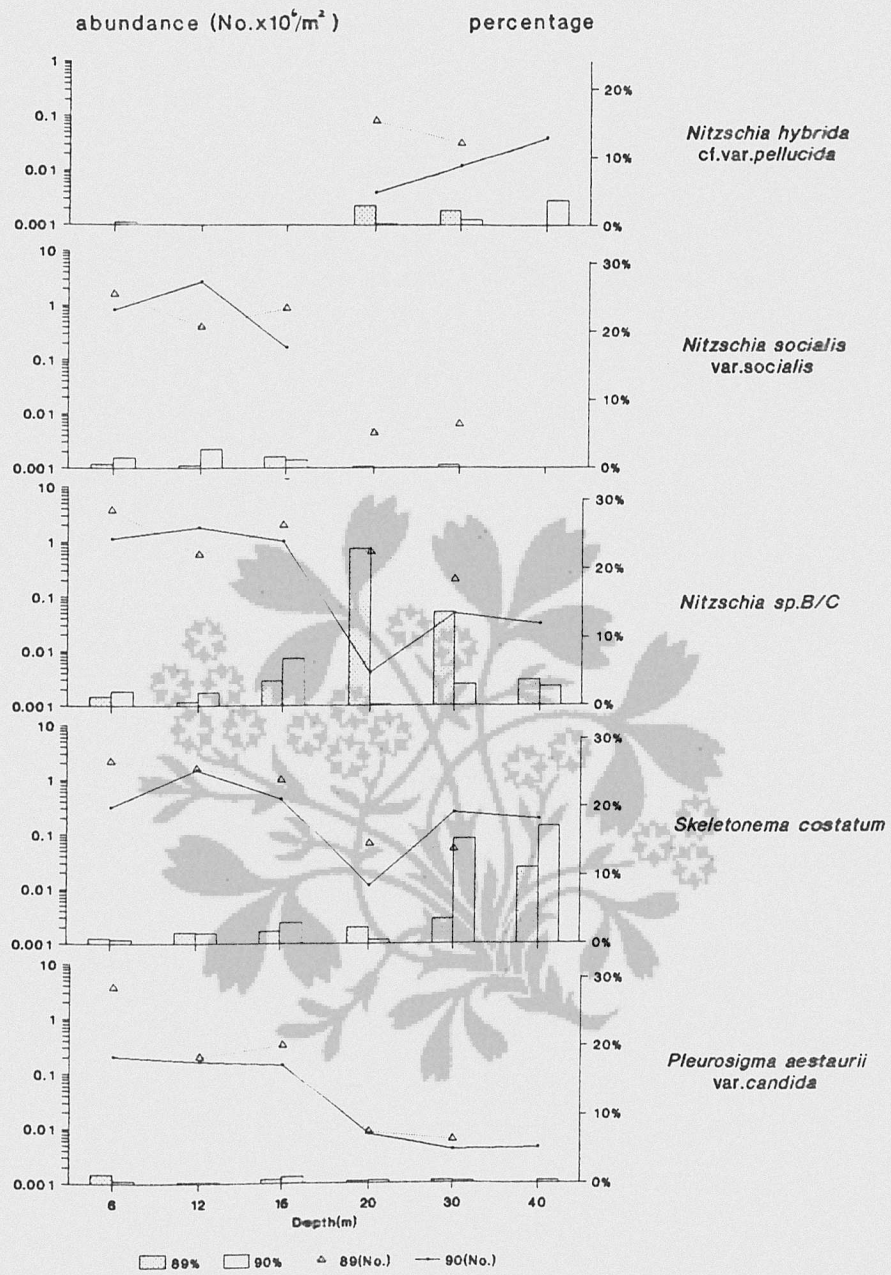
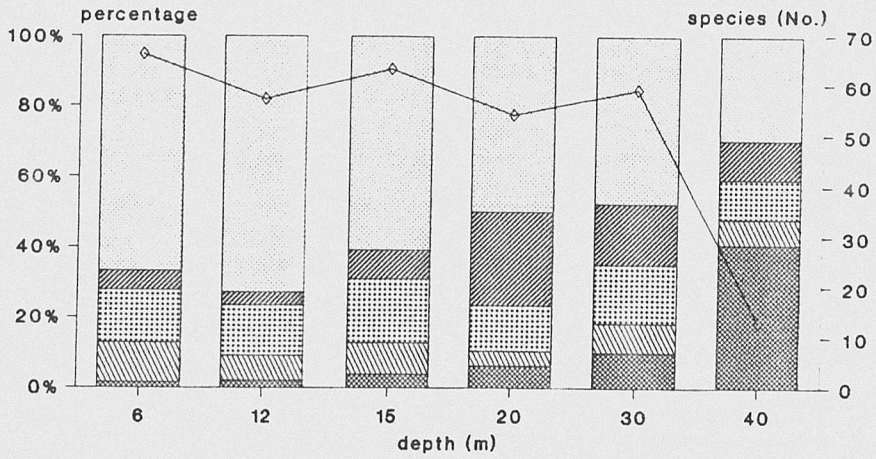
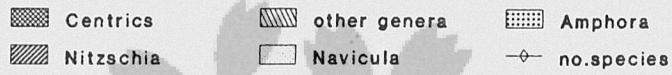


Fig.22. The depth distribution pattern of the predominant species of epipelagic diatom recorded from 6-40m depth in 1989 and 1990 indicated by estimated abundance.

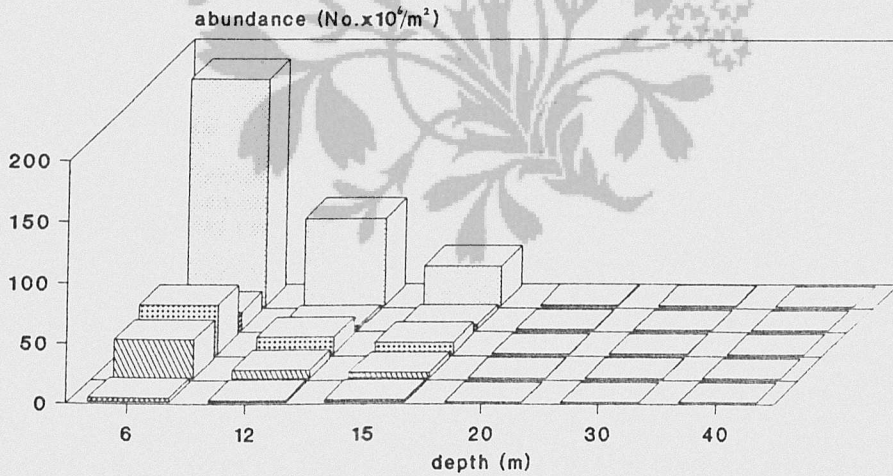
1989



A



1989



B

Fig. 23. The spatial change in the epipelagic community structure along a depth gradient (6-40m) in 1989.
 A: Estimated percentage occurrence of the major epipelagic diatom genera and number of species.
 B: Estimated abundance of major genera.

Table 6. List of epipelagic species and their percentage abundance in sample communities recorded along a bathymetric gradient (6 - 40m), within Loch Goil, in June 1989. Depth is in metres below chart datum. Key: * = rare species (see section 4.3.2.2).

Taxa	1989:	6m	12m	15m	20m	30m	40m
<i>Acnantes</i> cf. <i>angustata</i>		*			*	*	
<i>Achnantes</i> <i>conspicua</i> var. <i>conspicua</i>		*				*	
<i>Achnantes</i> <i>fimbriata</i>		0.4				0.5	
<i>Achnantes</i> sp.		0.4				0.5	
<i>Amphiprora</i> <i>plicata</i>		*	0.4				
<i>Amphora</i> cf. <i>arcus</i>		*	0.4		*		
<i>Amphora</i> <i>coffeaeformis</i> var. <i>acutiuscula</i>		3.0	0.6	4.9	*	0.5	*
<i>Amphora</i> <i>laevis</i> var. <i>perminuta</i>		0.8	*	*	0.3	*	
<i>Amphora</i> <i>ostrearia</i> var. <i>ostrearia</i>		0.4		0.4	*	0.7	
<i>Amphora</i> cf. <i>proteus</i> var. B		2.2	4.4	5.5	8.5	8.1	9.5
<i>Amphora</i> <i>proteus</i> var. <i>oculata</i>		1.2	2.9	1.3	1.7	2.5	
<i>Amphora</i> <i>proteus</i> var. <i>proteus</i>			0.4	*	*		
<i>Amphora</i> <i>truncata</i>		1.0	1.5	0.9	1.1	1.6	
<i>Amphora</i> <i>ventricosa</i>		3.6	4.0	5.3	1.3	1.4	
<i>Amphora</i> sp. A		0.4	*			*	
<i>Amphora</i> sp. B		2.4	*	0.4	*	2.5	
<i>Amphora</i> sp. C		*	*				
<i>Caloneis</i> <i>liber</i> var. <i>elongata</i>			*	*	*		
<i>Caloneis</i> <i>liber</i> var. <i>excentrica</i>		*					
<i>Campylodiscus</i> <i>fastuosa</i>			*	*			
<i>Chaetoceros</i> sp.							
<i>Cocconeis</i> <i>scutellum</i> cf. var. <i>scutellum</i>		*	0.4	0.4	*	*	*
<i>Coscinodiscus</i> sp.		0.6	0.4	2.1	3.9	6.8	38.1
<i>Cylindrotheca</i> <i>gracilis</i>							
<i>Cylindrotheca</i> <i>closterium</i>		1.4	*	1.5	*	0.5	
<i>Dictyocha</i> <i>speculum</i> var. <i>speculum</i>		*	*	*	*	*	*
<i>Diploneis</i> <i>aesteva</i> var. <i>delicata</i>		*	*	*	*	*	*
<i>Diploneis</i> <i>aesteva</i> var. <i>fusca</i>		*	*	*	*	*	*
<i>Diploneis</i> <i>crabro</i>		*	*	*	*	*	*
<i>Diploneis</i> <i>ovalis</i>		*	0.6	*	*	0.9	
<i>Diploneis</i> <i>smithii</i> var. <i>smithii</i>		0.4	*	0.4		0.5	
<i>Diploneis</i> <i>splendida</i>			*	*		*	
<i>Donkinia</i> <i>reticulata</i>		0.6	*		*	*	
<i>Eunotia</i> cf. <i>septentrionalis</i>				*	*		
<i>Eunotia</i> sp.		*					
<i>Frustulia</i> <i>rhomboides</i> var. <i>rhomboides</i>							
<i>Frustulia</i> <i>rhomboides</i> var. <i>saxonica</i> f.u						*	
<i>Gomphonema</i> <i>parvulum</i> var. <i>exilis</i>							
<i>Gyrosigma</i> <i>algoris</i>		0.4	0.4		0.6	0.5	
<i>Gyrosigma</i> <i>distortum</i> var. <i>parkeri</i>		*		*			
<i>Gyrosigma</i> <i>rectum</i>							
<i>Gyrosigma</i> <i>tenuirostrum</i>		*		*			
<i>Navicula</i> <i>abrupta</i>							
<i>Navicula</i> <i>arenaria</i>		1.6	2.9	7.5	14.1	9.3	9.5
<i>Navicula</i> <i>atlantica</i>				0.4		0.5	
<i>Navicula</i> cf. <i>bryophila</i>				0.4		*	
<i>Navicula</i> cf. <i>decussis</i> var. <i>decussis</i>							
<i>Navicula</i> cf. <i>delognei</i>							
<i>Navicula</i> <i>crucigera</i>		*	0.4	*			
<i>Navicula</i> <i>cryptocephala</i> * var. <i>veneta</i> (cf)*		3.6	*	3.0	0.6	1.1	
<i>Navicula</i> cf. <i>digito-radiata</i> var. B							
<i>Navicula</i> cf. <i>digito-radiata</i>							
<i>Navicula</i> <i>digito-radiata</i> var. <i>cyprinus</i>				*			
<i>Navicula</i> <i>distans</i>		2.6	2.7	2.1	0.9	3.2	

Continued....

Taxa	1989:	6m	12m	15m	20m	30m	40m	n
<i>Navicula forcipata</i> var. <i>densistriata</i>		7.3	7.8	5.5	7.1	9.7	9.5	
<i>Navicula gregaria</i>		*		0.6	0.5			
<i>Navicula hennedyi</i> var. <i>hennedyi</i>			0.4		*	*		
<i>Navicula palpebralis</i> var. <i>angulosa</i>		0.4		*	0.3	0.5		
<i>Navicula phyllepta</i>		1.0	*	0.4	*	1.1		
<i>Navicula plicata</i> var. <i>constricta</i>						*		
<i>Navicula retusa</i> var. <i>cancellata</i>								
<i>Navicula retusa</i> var. <i>gregorii</i>		3.2	1.1	0.9	1.4	2.9	*	
<i>Navicula tripunctata</i> * (cf.)*		31.3	43.5	34.1	15.7	12.4	9.5	
<i>Navicula</i> sp. A		1.4	1.1	0.6	0.6	1.1		
<i>Navicula</i> sp. B.		2.0	0.4	0.6	3.1	2.3		
<i>Navicula</i> sp. C/D		1.8	3.6	0.6	1.1	0.5		
<i>Navicula</i> sp. F/G/H		13.7	12.2	7.0	6.1	4.7	*	
<i>Navicula</i> sp. I								
<i>Nitzschia angularis</i>		1.0	1.1	1.7	0.5	0.5	9.5	
<i>Nitzschia angularis</i> cf. var. <i>affinis</i>		*						
<i>Nitzschia bilobata</i> var. <i>bilobata</i>				*				
<i>Nitzschia aff. distans</i>		*	*	*				
<i>Nitzschia hybrida</i> cf. var. <i>pellucida</i>					3.0	2.3		
<i>Nitzschia insignis</i> var. <i>smithii</i>		0.4	0.4	*		*		
<i>Nitzschia marginulata</i> var. <i>marginulata</i>		0.6						
<i>Nitzschia panduriformis/punctata</i>		0.6	0.6	*	0.3	0.5		
<i>Nitzschia sigma</i> var. <i>intercedens</i>		0.8	0.4	0.6				
<i>Nitzschia socialis</i> var. <i>socialis</i>		0.6	0.4	1.7	*	0.5		
<i>Nitzschia</i> sp. B/C		1.4	0.6	3.8	24.0	14.4	*	
<i>Opephora martyi</i>		*	*	*	*			
<i>Pinnularia ambigua</i>								
<i>Pinnularia appendiculata</i>					*			
<i>Pinnularia divergens</i>							*	
<i>Pinnularia rupestris</i>							*	
<i>Pinnularia</i> sp. A							*	
<i>Plagiogramma staurophorum</i>					*			
<i>Plagiotropis tayrecta</i>		*		*	*			
<i>Pleurosigma aestaurii</i> var. <i>candida</i>		1.8	0.4	1.5	0.3	0.5		
<i>Pleurosigma australe</i>			0.6	*		*		
<i>Pleurosigma formosum</i>		*	*	*	*			
<i>Pleurosigma naviculaceum</i>			*	*				
<i>Pleurosigma nubecula</i>								
<i>Pleurosigma salinarum</i>			0.4	0.4	*	*		
<i>Pleurosigma strigosum</i>		0.6	0.6	0.6	*	0.5		
<i>Rhabdonema minutum</i>								
<i>Rhoicosphenia abbreviata</i>			*					
<i>Skeletonema costatum</i>		0.8	1.7	1.9	2.5	3.8	14.3	
<i>Stauroneis salina</i> var. <i>lagerstedtii</i>		*	*	*	*	0.9		
<i>Stauroneis</i> sp.								
<i>Stenoneis inconspicua</i>		*						
<i>Stenoneis</i> sp.		1.6		*				
<i>Surirella fastuosa</i> var. <i>fastuosa</i>		*	*	*	*			
<i>Surirella hybrida</i>						*		
<i>Synedra</i> cf. <i>tabulata</i>		*			*			
<i>Tabellaria flocculosa</i>			*	*		*		
<i>Trachyneis aspera</i> var. <i>aspera</i>		*	*	*	0.3	*		
<i>Tropidoneis lepidoptera</i>					*	*		
<i>Tropinoneis vitrea</i> var. <i>scaligera</i>		0.4		0.4				
Number of species:		66	58	63	54	59	13	
Number of cells identified:		496	476	469	637	443	27	

Table 7. List of epipelagic species and their percentage abundance in sample communities recorded along a bathymetric gradient (6 - 40m), within Loch Goil, in July 1990. Depth is in metres below chart datum. Key: * = rare species (see section 4.3.2.2).

Taxa	1990:	6m	12m	15m	20m	30m	40m
Acnantes cf.angustata							
Achnantes conspicua var.conspicua							
Achnantes fimbriata			*	*			
Achnantes sp.				*	0.4	*	
Amphiprora plicata		*	*				
Amphora cf.arcus							
Amphora coffeaeformis var.acutiuscula	4.3		1.1	0.8		1.6	*
Amphora laevis var.perminuta	*				*	*	*
Amphora ostrearia var.ostrearia	0.6		*				
Amphora cf.proteus var.B	10.2		3.0	4.1	12.0	9.6	6.4
Amphora proteus var.oculata	2.0		2.2	3.6	2.4	1.6	1.3
Amphora proteus var.proteus	0.6		*	*		*	*
Amphora truncata	0.8		*	*	0.6	0.8	*
Amphora ventricosa	2.3		3.9	1.0	0.8	1.6	*
Amphora sp.A							
Amphora sp.B	*						
Amphora sp.C						0.8	
Caloneis liber var.elongata							
Caloneis liber var.excentrica							
Campylodiscus fastuosa			*				
Chaetoceros sp.			*	0.5	0.8	2.4	2.1
Cocconeis scutellum cf.var.scutellum	0.6			*	0.4	0.5	1.7
Coscinodiscus sp.	1.4		2.2	7.1	18.4	18.9	21.0
Cylindrotheca gracilis	*					*	
Cylindrotheca closterium	*			*	*	*	
Dictyocha speculum var.speculum					*	0.5	
Diploneis aesteva var.delicata	*		*				
Diploneis aesteva var.fusca	*		*		*		
Diploneis crabro							
Diploneis ovalis	*		*			*	*
Diploneis smithii var.smithii	*		0.4		*	*	
Diploneis splendida			*				
Donkinia reticulata					*	*	*
Eunotia cf.septentrionalis	0.4		*		0.4		
Eunotia sp.							*
Frustulia rhomboides var.rhomboides						*	
Frustulia rhomboides var.saxonica f.u							
Gomphonema parvulum var.exilis							*
Gyrosigma algoris	*		*	0.3			
Gyrosigma distortum var.parkeri							
Gyrosigma rectum					*		
Gyrosigma tenuirostrum							
Navicula abrupta				*			
Navicula arenaria	2.7		6.1	15.8	11.0	4.5	5.2
Navicula atlantica							
Navicula cf.bryophila					0.4	*	
Navicula cf.decussis var.decussis						0.5	
Navicula cf.delognei	*						
Navicula crucigera							
Navicula cryptocephala* var.veneta (cf)*	2.2		*	1.3	1.6	1.3	3.9
Navicula cf.digito-radiata var.B						0.5	
Navicula cf.digito-radiata			*				
Navicula digito-radiata var.cyprinus							
Navicula distans	5.3		0.6	3.5	2.4	3.5	1.7

Continued....

Taxa	1990:	6m	12m	15m	20m	30m	40m
<i>Navicula forcipata</i> var. <i>densistriata</i>		8.6	19.2	9.5	13.4	7.7	5.2
<i>Navicula gregaria</i>						*	
<i>Navicula hennedyi</i> var. <i>hennedyi</i>			*				
<i>Navicula palpebralis</i> var. <i>angulosa</i>		*	*	*	0.4		
<i>Navicula phyllepta</i>		0.4	*				*
<i>Navicula plicata</i> var. <i>constricta</i>			*			*	
<i>Navicula retusa</i> var. <i>cancellata</i>							*
<i>Navicula retusa</i> var. <i>gregorii</i>		0.8		0.7	*		
<i>Navicula tripunctata</i> * (cf)*		27.2	44.5	32.9	25.7	13.6	15.0
<i>Navicula</i> sp. A		*	*	*	*	*	
<i>Navicula</i> sp. B.		1.4	0.9	1.3	2.6	1.1	3.0
<i>Navicula</i> sp. C/D		0.4	1.3	1.8	1.0	1.3	
<i>Navicula</i> sp. F/G/H		21.1	6.8	2.3	3.4	6.1	4.3
<i>Navicula</i> sp. I							*
<i>Nitzschia angularis</i>		0.4	0.4	0.5			*
<i>Nitzschia angularis</i> cf. var. <i>affinis</i>		1.0		*	*	0.8	3.0
<i>Nitzschia bilobata</i> var. <i>bilobata</i>							
<i>Nitzschia</i> aff. <i>distans</i>		*	*				
<i>Nitzschia hybrida</i> cf. var. <i>pellucida</i>		0.4			*	0.8	3.9
<i>Nitzschia insignis</i> var. <i>smithii</i>							
<i>Nitzschia marginulata</i> var. <i>marginulata</i>							
<i>Nitzschia panduriformis/punctata</i>		0.4	*	*	0.6	*	
<i>Nitzschia sigma</i> var. <i>intercedens</i>		*	0.4				
<i>Nitzschia socialis</i> var. <i>socialis</i>		1.6	3.0	1.2			
<i>Nitzschia</i> sp. B/C		2.2	2.0	7.1	*	3.2	3.0
<i>Opephora martyi</i>			*				*
<i>Pinnularia ambigua</i>		*					
<i>Pinnularia appendiculata</i>				*	*	*	
<i>Pinnularia divergens</i>							
<i>Pinnularia rupestris</i>							
<i>Pinnularia</i> sp. A							
<i>Plagiogramma staurophorum</i>							
<i>Plagiotropis tayrecta</i>		*					
<i>Pleurosigma aestaurii</i> var. <i>candida</i>		0.4	*	1.0	0.4	*	*
<i>Pleurosigma australe</i>			*				
<i>Pleurosigma formosum</i>							
<i>Pleurosigma naviculaceum</i>					*		
<i>Pleurosigma nubecula</i>		*					
<i>Pleurosigma salinarum</i>			*			0.5	
<i>Pleurosigma strigosum</i>			0.4	0.3	0.4		0.9
<i>Rhabdonema minutum</i>		*					*
<i>Rhoicosphenia abbreviata</i>					*		*
<i>Skeletonema costatum</i>		0.6	1.7	3.1	0.6	16.0	18.5
<i>Stauroneis salina</i> var. <i>lagerstedtii</i>							
<i>Stauroneis</i> sp.					*		
<i>Stenoneis inconspicua</i>							
<i>Stenoneis</i> sp.							
<i>Surirella fastuosa</i> var. <i>fastuosa</i>							
<i>Surirella hybrida</i>							
<i>Synedra</i> cf. <i>tabulata</i>			*				
<i>Tabellaria flocculosa</i>			*	0.3		*	
<i>Trachyneis aspera</i> var. <i>aspera</i>		*					
<i>Tropidoneis lepidoptera</i>			*		*		
<i>Tropidoneis vitrea</i> var. <i>scaligera</i>							
Number of species:		47	48	35	40	42	34
Number of cells identified:		511	541	608	501	375	233

	1989	abundance	chlorophyll-a	carotenoids	salinity	temperature	oxygen	depth	axis 1	axis 2	silt (%)	median
abundance		N
chlorophyll-a		.	P
carotenoids	
salinity	
temperature	
oxygen	
depth	
axis 1	
axis 2	
silt (%)	
median	
	1990	abundance	chlorophyll-a	carotenoids	salinity	temperature	oxygen	depth	axis 1	axis 2	silt (%)	median
abundance		P	P	.	.	P	.	N	N	.	.	.
chlorophyll-a		.	P	N	.	.	.
carotenoids		P	.	N
salinity		N
temperature		N	.	.	.
oxygen	
depth	
axis 1	
axis 2	
silt (%)	
median		P

Table 8. The relationships between scores on DECORANA axis with epipelagic abundance, chlorophyll-a and environmental variables. Data was recorded along a bathymetric gradient in 1989 and 1990. Significant levels ($P < 0.050$) derived from Spearman rank correlation analysis. Median = median sediment particle size; '.' = no correlation; P = positive correlation; N = negative correlation.

major taxa (Fig. 23) and most of the rarer species (Fig. 22).

4.3.3.2.2 July 1990

TWINSpan analysis of the 1990 community data separates the shallow water stations (6m, 12m, 15m) from the deeper stations (20m, 30m, 40m) in a manner similar to that seen in June 1989 (Fig. 24, 1990). However, examination of both the dendrogram (Fig. 19, 1990) and ordination plot (Fig. 20; 1990) shows that the shallow community is less distinct than that recorded in the previous year with a lower level of similarity (cluster analysis) and a greater spread along the principal ordination axis (eigenvalue = 0.221, 87.35 % of the variability). DECORANA and cluster analyses also indicate a difference between the community of the two deeper stations (30m and 40m) and the epipelon recorded at the shallower stations. This can be attributed to a marked increase in abundance of *Skeletonema costatum* at these two stations (Fig. 20).

Examination of the TWINSpan matrix (Fig. 24, 1990) and Table 7 once again reveals the reason for this grouping, with several of the less abundant taxa showing highly restricted distribution along the bathymetric gradient. In particular, four of the taxa that characterised the shallow community in 1990 (*Amphora plicata*, *Nitzschia distans*, *Nitzschia sigma* var. *intercedens* and *Campylodiscus fastuosus*) were also limited to the same range of depth in 1989. However, of those species that characterised the two deepest stations (20m and 40m), only *Frustulia rhomboides* was common to both 1989 and 1990 communities. *F. rhomboides* is strictly a fresh water species which would have had to have been washed in from Cormonachan Burn. None the less the cell was alive within the sediment which suggests that I may have identified this species incorrectly (see appendix 1). The taxa that separate the deep and shallow water communities in 1989 and 1990 bear little resemblance;

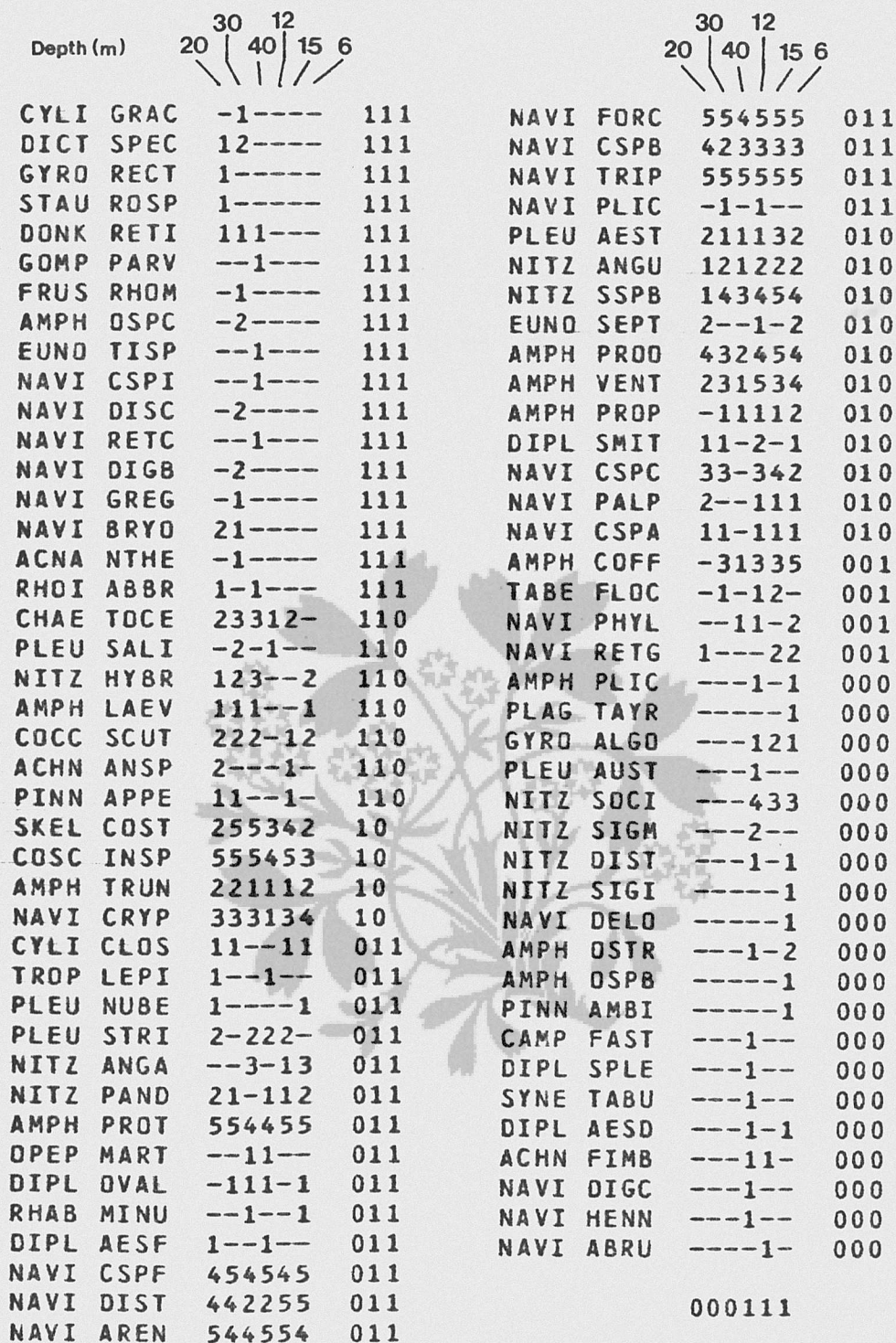
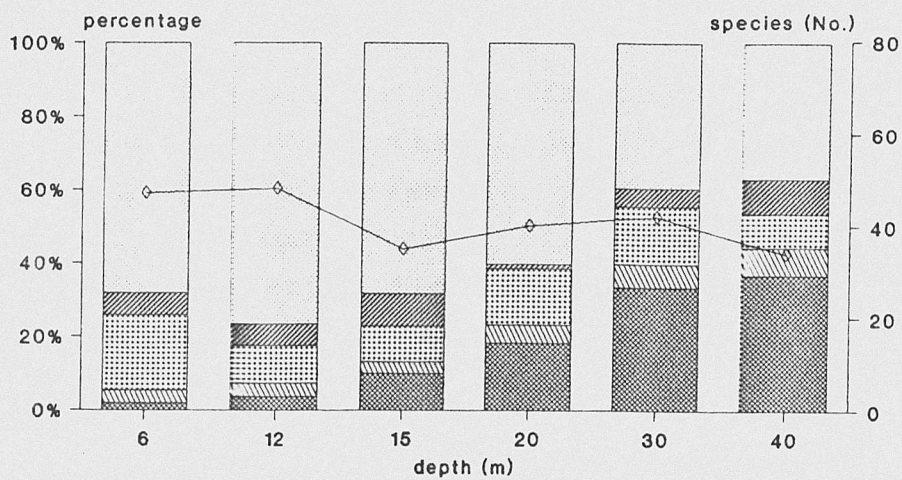


Fig. 24.

1990: TWINSpan classification of epiplic communities recorded in Loch Goil at a range of depths in 1990. Abbreviations of species names are shown at the left; sample numbers along the top. The classification of species and samples are indicated along the right and bottom margins (Hill, 1979 [a]). The main dichotomy for the samples is indicated by a vertical separation. Values indicate a scale of abundance, with absence of a species represented by the symbol "-".

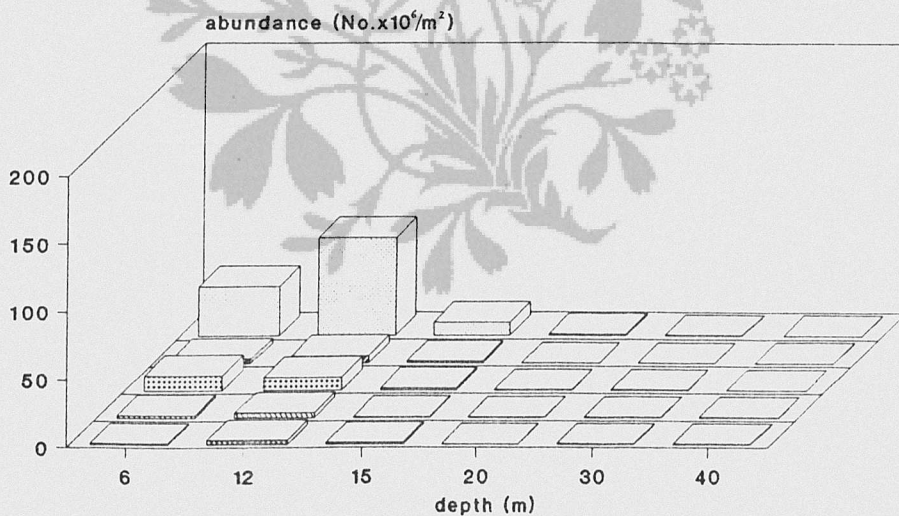
1990



A

Centrics other genera Amphora
 Nitzschia Navicula species

1990



B

Fig. 25. The spatial change in the epipellic community structure along a depth gradient (6-40m) in 1990.
 A: Estimated percentage occurrence of the major epipellic diatom genera and number of species.
 B: Estimated abundance of major genera.

however, it is the re-occurrence of the most numerically dominant taxa that is most notable feature of the data. Inspection of Fig. 22. shows the similarity, between the two years, of the bathymetric distribution of the major taxa (*Navicula cf.tripunctata*, *Navicula forcipata* var. *densistriata*, *Navicula arenaria*, *Navicula* sp.F/G/H, and *Amphora cf. proteus* var. B). This similarity is summarised by comparing Fig. 23, (A) and Fig. 25, (A) which divide the sample communities of the 1989 and 1990 data (respectively) into the relative proportions of the major genera.

The most notable differences in the distribution and abundance of the major taxa, between the two years, is the absence of a peak (recorded in 1989 at 20m and 30m depth) of *Nitzschia* sp. B/C and an increase in the abundance of *Coscinodiscus* sp. at 20m, 30m and 40m in 1990. Fig. 22 and Fig. 25 shows that it was an increase in the abundance of the dominant *Navicula* and *Amphora* species (*Navicula cf.tripunctata*, *Navicula forcipata* var. *densistriata*, *Navicula arenaria*, *Amphora ventricosa* and *Amphora proteus* var. *oculata*) at a depth of 12m that was responsible for the peak in algal density recorded at this depth (Fig. 18).

4.3.3.3 Physicochemical Gradients.

Changes in the temperature (Fig. 26) and oxygen levels (Fig. 27) with depth in both 1989 and 1990 show that the sampled habitats were within the epilimnion of Loch Goil with no evidence of stratification within the measured depth range (6-40m). In 1989 temperature (9-8.4 °C) and oxygen (8.6-7.5 ppm) showed little change as depth increased indicating a well mixed water column. In 1990 temperature (12.5-8.8 °C) and oxygen (12.2-5.5 ppm) showed greater variation with both parameters decreasing in an approximately linear fashion with increasing depth. The variation in salinity with depth can be seen in Fig.28. However, these data appear to be erroneous as levels differ considerably from those recorded from the centre of Loch Goil by the Clyde River Purification Board (CRPB) in

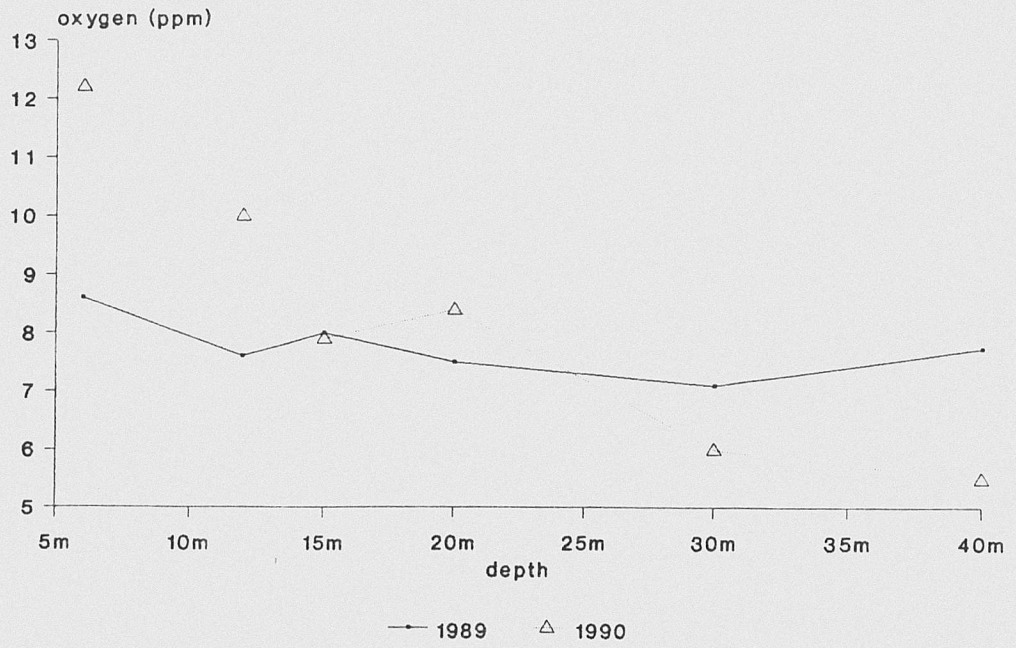


Fig.26. Spatial distribution of oxygen (at sediment/sea water interface) along a depth gradient.

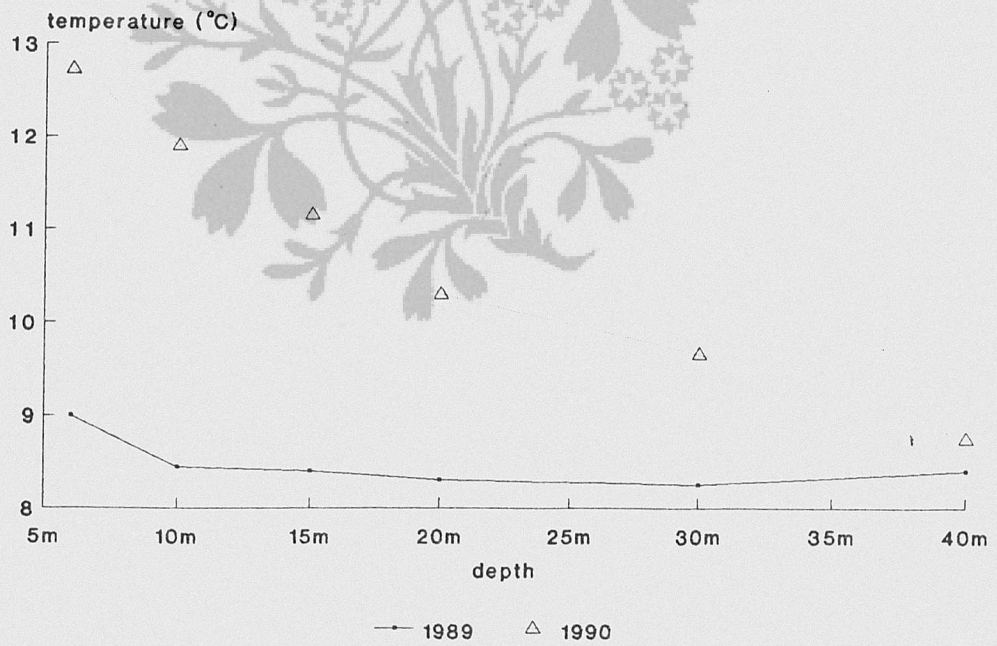


Fig.27. Spatial distribution of temperature at the sediment surface along a depth gradient.

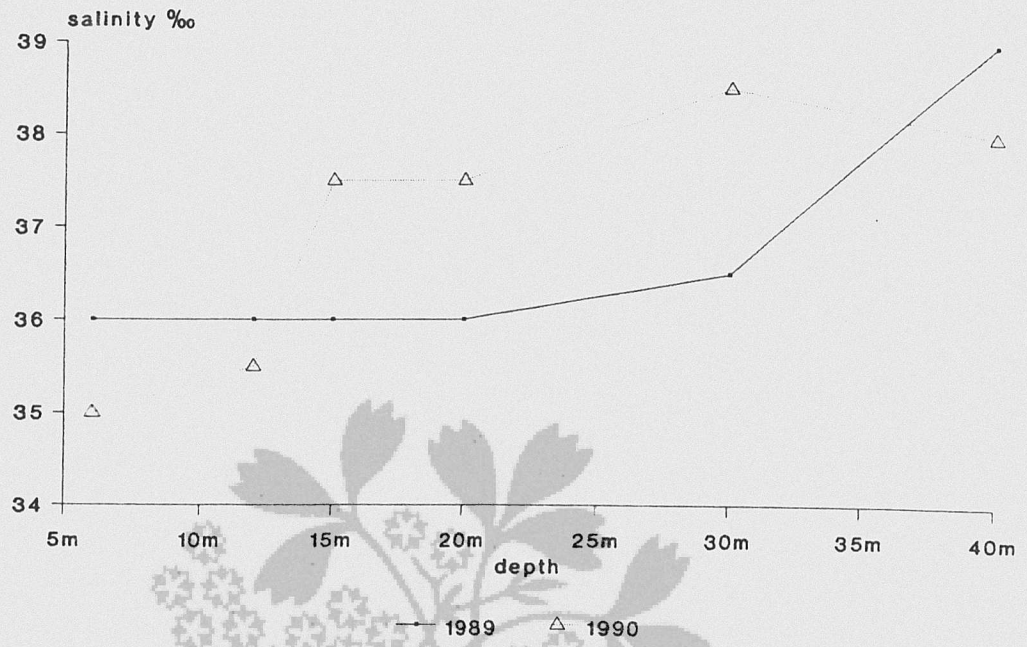


Fig.28. The salinity of seawater (recorded at the sediment surface) along a depth gradient.

depth (m)	%<63 silt	%>63u sand	silt:sand	(phi) mean	(phi) medi	(phi) mode	sorting
1989							
6m	17.1	82.9	0.21	3.40	3.45	4.00	0.80
12m	12.9	87.1	0.15	3.18	3.25	4.00	0.90
15m	5.2	94.8	0.05	1.16	1.95	-3.00	1.55
20m	35.7	64.3	0.56	5.30	3.75	4.00	1.40
30m	25.7	74.3	0.35	3.47	3.60	4.00	1.23
40m	23.0	77.0	0.30	3.56	3.50	4.00	1.12
1990							
6m	47.8	52.2	0.91	3.37	3.85	-1.75	2.83
12m	24.8	75.2	0.33	3.61	3.50	3.95	1.18
15m	22.0	78.0	0.28	3.44	3.35	3.95	1.08
20m	25.8	74.2	0.35	4.29	3.10	3.95	2.33
30m	26.0	74.0	0.35	1.98	2.90	-1.75	3.48
40m	58.6	41.4	1.41	4.68	4.45	3.95	1.95

Table 9. Sediment particle size data recorded along a depth profile (6 - 40 m) of Loch Goil in June 1989 and July 1990.

July 1989 [1m = 30.91 ppt; 40m = 32.77 ppt] and July 1990 [1m = 30.48 ppt; 40m = 32.98 ppt] (Fig. 29; CRPB, unpubl. data). Although the magnitude of the data appear to be incorrect, if the errors are consistent then a halocline existed at a depth of 12-15m in 1990.

Although no data were obtained showing light attenuation, the light available to the epipelon appeared to be limited in both 1989 and 1990 by a phytoplankton bloom. This formed a dense layer that discoloured the water green from the surface to a depth of approximately 2-5m. In July 1990 phytoplankton density in the surface water was particularly dense, causing very poor (< 0.5m) visibility in the first few meters of water; compared with approximately 2m surface visibility in 1989.

Spearman Rank analysis of the 1989 data revealed a significant (negative) correlation ($P < 0.05$) between epipellic abundance and depth. However, no other associations were found between either axis 1 or abundance and any environmental variables (Table 8). In 1990 significant correlations ($P < 0.05$) were found between axis 1 (DECORANA analysis) and both depth (positive) and temperature (negative). Similarly, the abundance of the epipelon was correlated ($P < 0.05$) with both depth (negative) and temperature (positive).

A cursory inspection of Loch Goil suggested a transition from a coarse sand in the shallows to finer muddy sediment with increasing depth. However, Table 9 shows there to be a mixed, heterogeneous distribution of sediments with no clear pattern of variation along the bathymetric gradient in either 1989 or 1990. Spearman rank analysis showed there to be no significant relationship between the epipelon and either the proportion of silt within the sediment or median sediment particle size (Table 8).

4.3.3.4 Phytoplankton

In June 1989 the predominant species found in the water column was *Chaetoceros* spp. Other phytoplankton

species present (approx. equal proportions) included: *Skeletonema costatum*, *Coscinodiscus* sp, *Diplopeltopsis minor*, *Noctiluca scintillans*, *Protooperidinium* sp, *Gonyaulax* sp, and euglenoid flagellates.

In July 1990 the turbid, green discolouration of the upper 6m of the water column was due to a bloom of the coccolithophore *Emiliana huxleyi*. Other species of phytoplankton present (approx. equal proportions) included: *Dictyocha speculum*, *Chaetoceros* spp, *Thalassiosira* sp, *Ceratium fusus*, *Gonyaulax* sp and euglenoid flagellates.

4.3.4 Discussion

The epipelon showed considerable variation in community composition, abundance and biomass along the depth gradient in both 1989 and 1990. The epipelon can be characterised into shallow and deep water zones, with differences in the density of algae and composition of the rarer species along the depth gradient.

The data identify a number of taxa that appear to inhabit a restricted range of depth but only four of these species were common to the shallow water community in both 1989 and 1990. Novel species occur in both years and some species (eg. *Nitzschia socialis*) that characterised the shallow zone in one year, were recorded at greater depth or at a range of depths in the other year. The selectivity of taxa for a particular depth has been observed by previous investigators in freshwater (Round, 1961; Roberts and Boylen, 1988; Kingston *et al*, 1983; Stevenson and Stoermer, 1981; Stevenson *et al*, 1985.) and marine (Bodeanu, 1964; Manea and Skolka, 1961) habitats. None of the six species of diatom recorded by Manea and Skolka (1961) in the Roumanian Black Sea were found in Loch Goil. However, 13 of the 86 taxa recorded by Bodeanu (1964) were recorded along the bathymetric gradient of Loch Goil. In particular, *Nitzschia distans*, *Nitzschia sigma* (4-10m) and *Nitzschia bilobata* (4-16m) were found by Bodeanu, and

myself, to be restricted to a similar depth range within the shallow sublittoral region. Most of the other species common to both studies (*Navicula forcipata*, *Navicula cf. cryptocephala*, *Amphora proteus*, *Amphora truncata*, *Amphora coffeaeformis* var. *acutiuscula*, *Nitzschia hybrida*, *Surirella fastuosa*, *Cocconeis scutellum* and *Diploneis smithii*) were more cosmopolitan in distribution; recorded by Bodeanu over most of the recorded depth range (2-20m). This is consistent with my data as these species were recorded over a similar range of depth (6-20m) within Loch Goil. The only exception is *Navicula retusa* var. *cancellata*, which was only recorded in Loch Goil at a depth of 40m, but was found at 4-16m depth in the Black Sea study.

An interesting feature of the data is the dominance within the sample communities (1989 and 1990) of the following taxa: *Navicula cf. tripunctata*, *Navicula forcipata* var. *densistriata*, *Navicula distans*, *Navicula arenaria*, *Amphora cf. proteus* var. B and *Navicula* sp. F/G/H (No distinction was made between *Navicula* sp. F, G or H in the analysis). These dominant species are different from the major community (see 4.3.1) recorded in Loch Sween over a 1-20m depth range by Smyth (1955). However, comparison of the whole of both communities shows there to be a number of common species: *Amphiprora* sp., *Amphora coffeaeformis*, *A. laevis*, *A. proteus*, *A. truncata*, *Cocconeis scutellum*, *Diploneis* sp., *Navicula abrupta*, *N. cryptocephala*, *N. digito-radiata*, *N. forcipata*, *N. gregoria*, *N. palpebralis*, *N. retusa*, *Nitzschia sigma*, *Pleurosigma formosum*, *Rhabdonema minutum*, *Tropidoneis* sp. Although the algal dominants may differ, the proportions of the dominant algae remain relatively unchanged over this depth range in both Loch Goil (6-30m) and Loch Sween (1-20m). However, the data collected by Smyth may not reflect the natural community present within the sediment as the methods were qualitative. In addition, because the slides were removed by lifting the exposed sample through the

water column (by a cord), the sample communities are likely to have been modified through the loss of algae.

The stability of the epipellic community in Loch Goil with depth differs from the communities recorded in the Black Sea (Bodeanu, 1964). With the exception of *Amphora coffeaeformis* var. *acutiuscula*, which was found to be the most abundant taxa over the 4-12m depth range, Bodeanu generally found a shift in the major taxa with increasing depth.

Comparison of the Loch Goil epipelon with freshwater communities (Round, 1961; Roberts and Boylen, 1988; Kingston et al, 1983; Stevenson and Stoermer, 1981; Stevenson et al, 1985.) shows there to be (not surprisingly) an entirely different range of species in the marine and freshwater habitats. However, the epipellic algal community recorded by Stevenson and Stoerma (1981) was also dominated by diatoms at all recorded depths (6.5-27.4m) although the diversity of algae (255 taxa, 36 genera) and abundance of diatoms (9×10^9 cells m^{-2} to 3×10^{10} cells m^{-2}) was higher than levels recorded in Loch Goil. This is in contrast with the results of Roberts and Boylen (1988) who found epipellic diatoms to have the greatest abundance (3×10^9 cells m^{-2}) at 1m and 4m depth whereas blue-green algae were found to dominated the deepest (7m) part of the lakes.

The increase in the proportion of planktonic species (*Chaetoceros* sp, *Skeletonema costatum*, *Coscinodiscus* sp) within the deepest (30m and 40m) sample communities of Loch Goil has also been observed in fresh water lake communities (Kingston et al, 1983; Stevenson and Stoerma, 1981). This is attributed to the deposition of more plankton (and less epipelon) in proportion to the longer water column of deep water areas.

In relation to the density of epipellic algae along the bathymetric gradient: the shallow zone (6-15m) was characterised in both 1989 and 1990 by relatively high epipellic abundance and pigment levels (chlorophyll-a and

carotenoids). The levels of algae were generally higher in 1989, particularly at the shallowest station (6m) where over five times more algae were recorded than the same depth in 1990. It was the shallowest depth in 1990 that possessed the highest algal density whereas a peak in density and biomass was recorded at a depth of 12m in 1990. The reason for the peak in epipellic pigment concentration at 12m depth in 1989, while cell numbers were proportionally lower, is not clear. However, *Navicula cf. tripunctata*, which formed 43.5 % of sample community at this depth, shows considerable size variation (24-67µm; see Appendix 1) and may have been generally larger at this depth. Further analysis of the sample communities would be required to test this hypothesis.

Round (1981) indicates that optimum standing crop of benthic algae does not always correspond to the shallowest depth, and the density of epipellic in fresh water lakes (Roberts and Boylen, 1988 and Stevenson & Stoermer, 1981) have been shown to peak at sub-surface depths of 9.1m and 4m respectively. Plante-Cuny recorded a peak of cell numbers (approx. $150 \text{ cells m}^{-2} \times 10^8$) at 30-35m depth, considerably deeper and higher than the pattern recorded in Loch Goil. Plante-Cuny's detailed floristic study also recorded a greater diversity of algae at depth (eg. an average of 42 species at 40-60m depth) and revealed a few species of epipellic diatoms living at depths of 300m and one species, *Navicula pennata*, at 360m. The occurrence of epipellic diatoms in these deep oceanic sediments can probably be related to the clarity of the water off the coast of Marseille compared with the relatively turbid waters of Scottish sea lochs. Another possible explanation is that the epipellic algae existing at these great depths are apochlorotic (without chlorophyll); existing heterotrophically.

The approximately linear decrease in epipellic density with depth, observed in 1989, is consistent with the pattern of distribution observed by Bodeanu (1964)

although the density of algae in this study of the Black Sea is considerably greater (9×10^9 cells m^{-2} to 3×10^{10} cells m^{-2}). Similarly, Stevenson *et al* (1985) observed a negative, linear relationship between the biovolume of most epipelagic algal species with depth (1- $<20m$) in New Hampshire lakes.

This investigation has characterised the epipelon along a gradient of depth within Loch Goil. However, it does not conclusively show the cause of these patterns of variation. Spearman rank analysis of the physicochemical data revealed significant correlations ($P = <0.05$) between epipelagic abundance and temperature in 1990. However, in 1989 the water temperature varied only $0.5 \text{ }^\circ\text{C}$ over the depth gradient whereas the density and composition of the epipelon varied considerably with depth.

The negative correlation, observed in both 1989 and 1990, between epipelagic abundance and depth indicates that a positive correlation would exist between the epipelagic abundance and light levels. However, this assumes that water clarity is spatially constant within Loch Goil. Sundback (1986) has shown that:

"turbidity may decrease with increasing depth, and thus light intensity at the sediment surface at 15m is not necessarily less than that at 8-10m".

Although the measurement of vertical light attenuation was not successfully made (due to faults in the electrical circuits) it is generally acknowledged that the amount of light transmitted to the substratum decreases with increasing depth and the spectral quality of the light is modified as it passes through water (Kendrick and Kroenberg, 1986). Light attenuation with depth is likely to be the principal factor influencing the bathymetric distribution of epipelagic density. Benthic microalgae are known to attain light saturated photosynthesis at a range of light intensities and inhibition of photosynthesis has been recorded when light intensity is greater than $150 \text{ } \mu\text{E} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ whereas $20 \text{ } \mu\text{E} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ was the recorded lower limit for microphytobenthic

growth (Sundbäck, 1986). Photosynthetic inhibition is unlikely to be the cause of the relatively low algal density recorded at 6m in 1990, as reduced visibility within the shallows appeared to severely limit the amount of light penetrating beneath the surface of the loch. However, the masking of light by the dense planktonic bloom of *Emiliana huxleyi* may have reduced the density of epipelton recorded along the whole bathymetric gradient in July 1990, compared with levels found in June 1989. Two weeks later (1st. August 1990), the bloom of *Emiliana huxleyi* turned the waters of Loch Goil, Holy Loch and Loch Long, a turquoise colour and reduced visibility within Loch Goil to less than 50 cm (pers. observation). This attracted the attention of both the media and the Clyde River Purification Board who attributed the colour to the scattering of light by the myriad coccolith scales released from the senescent *Emiliana huxleyi* cells (C.R.P.B pers. comm.).

The changes in the epipellic community with depth may reflect the adaptation of some taxa to low light regimes. Rosen and Lowe (1984) have observed that some diatom species (eg. *Cyclotella meneghiniana*) adapt to the low light intensities that are present at depth, by increasing chlorophyll-a concentration, chloroplast volume and thylakoid surface density. Certain species of *Navicula* and *Nitzschia* have also been shown to be facultative heterotrophs (Stevenson *et al*, 1985, Admiraal and Peletier, 1979. To determine the importance of light as a determinant, future study should include experimental manipulation of natural cultures of epipelton to varied light intensities and wavelengths. Natural epipellic communities could also be manipulated in field experiments by transplanting shallow water communities to deeper areas and by modifying the light regime, *in situ*. Future study of the bathymetric distribution of the epipelton should include accurate measurement of light intensity. It should be noted, however, that the methods used will have to be

more sensitive than those routinely used in phytoplankton research, where the depth of the euphotic zone is assumed to correspond to the depth where 1% of the incident light remains.

Sundbäck (1986) also observed optimum levels of epipellic chlorophyll-a at 14-16m depth and related this to the depth of the halocline and the adaptation^{ta} of taxa, recorded at that depth, to low light intensities. This is comparable to my data, as the peak in epipellic density at 12m depth (July 1990) coincides with the depth of the halocline observed both in my data (Fig. 28) and in that recorded by the CRPB (Fig. 29; CRPB, unpubl. data). The CRPB data also shows there to be thermal stratification at 30-40m in July 1989 and 40-50m in July 1990 further indicating that the sampled depth range is within the epilimnion of Loch Goil. Epipelon were found with protoplasm in them from the deepest recorded depth (40m) in both 1989 and 1990. However, future studies should sample from a greater range of depth (0-93m), in both the summer and winter months, using an automatic corer such as a "Craib" corer (Mc.Intyre and Warwick, 1971), operated from a surface vessel with winch. This would ascertain whether summer stratification, and subsequent stagnation (oxygen <2 ppm) of the deep (30-50m) waters of Loch Goil (Edwards *et al*, 1986; Mackay and Halcrow, 1976; CRPB unpublished data) is reflected in changes in the community structure of the epipelon. It would also determine the maximum depth at which the epipelon inhabits Loch Goil. Round (1972) stipulates that "epipellic populations in large (fresh) water bodies tend to occur only above the thermocline". Further investigation is required to ascertain whether or not this is true for Loch Goil and sea loch habitats in general.

Other factors that may influence the bathymetric distribution of epipelon include: wave action, nutrient levels of interstitial water, organic content of the sediment, sediment redox potential, grazing pressure and

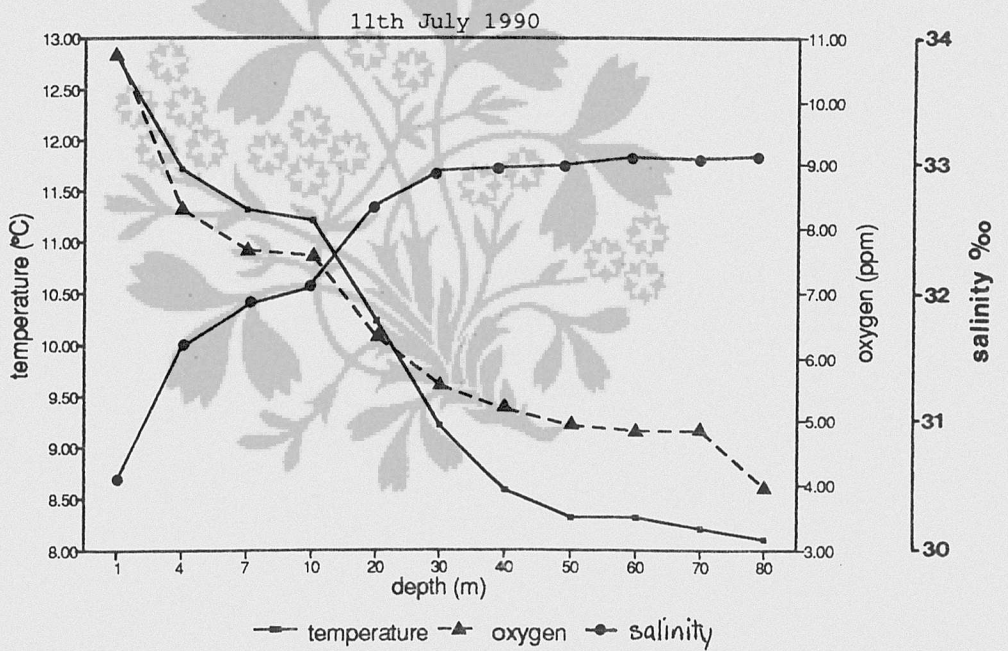
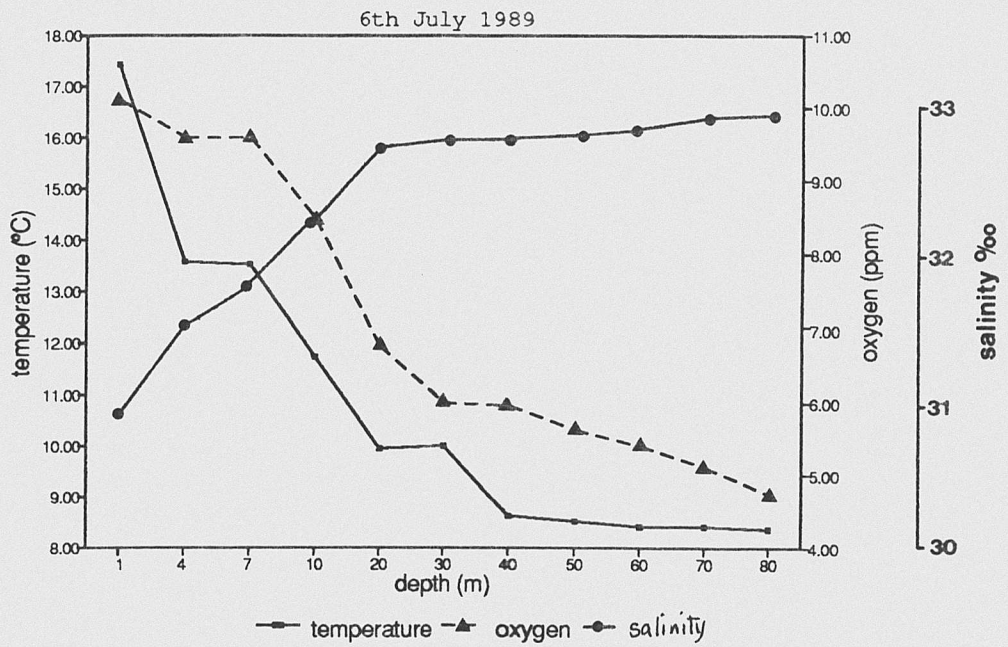


Fig. 29. Temperature, salinity and oxygen profiles along a bathymetric gradient (0-80m) in Loch Goil during July 1989 and July 1990 (CRPB unpublished data).

bioturbation. Wave action may disrupt surface communities in shallow water (Gruending, 1971; Stevenson and Stoermer, 1981). However the effect on the shallow water community in Loch Goil is unlikely to be significant as the site is extremely sheltered and the intensity of winds preceding these surveys were minimal. Sediment is generally considered to be an inexhaustible source of nutrients for the epipelon, however, nutrient limitation has been demonstrated (Graneli and Sundbäck, 1985) and availability of inorganic nutrients may effect species composition (Pringle and Bowers, 1984). A future study could include measurement of micronutrient concentrations (NH_3N , TON , $\text{PO}_4\text{-P}$, SiO_2) within interstitial water; sediment redox potentials and the organic content of the sediment at different depths. The subtidal epipelon may also be influenced by grazing and bioturbation of benthic macro-invertebrates along the depth gradient. Qualitative observations made while sampling showed there to be an extremely diverse and spatially heterogeneous community of particularly: polychaetes, ophiuroids, anthozoans, pennatulids, holothurians, and bivalves. The effect of these animals on the abundance and community composition of the epipelon requires investigation. In particular the characterisation of the invertebrate community along the same bathymetric gradient would indicate possible trophic relationships involving the epipelon (see section 5.1). This could lead to more detailed examination of grazing and bioturbation effects through experimental investigation in the field and laboratory.

This investigation has succeeded in characterising the bathymetric distribution of epipelon in Loch Goil, providing novel data on a habitat not previously described. Graphical representation of the data and the results of the multivariate analysis provides a visual display of variations in the epipelon which, in itself, is convincing evidence of environmental and biological control of the epipellic algae over the measured depth

gradient. The analyses have lead to the rejection of the null hypothesis (H_0): "there are no differences in the epipelton along a bathymetric gradient, in Loch Goil".

4.4.1 Introduction

There have been many studies reporting the seasonal succession of epipelic algae in freshwater habitats (Hickman, 1975; Round, 1972; Gruendling, 1971) and littoral marine habitats (Assus, 1982; Colijn and Dirksen, 1981; Colijn et al., 1975; Round, 1980; Hopkins, 1964; Castenholz, 1967; Admiraal et al., 1984). A number of authors also describe seasonal changes in epipelic standing crop in marine (eg. Jonge, 1980; Jonge and Colijn, 1982; Sayth, 1955) and freshwater (Moss, 1969; Hickman and Round, 1970) habitats. Perhaps it is because of sampling difficulties that data on the seasonality of epipelton in the epipelton are scarce in the literature. However, they will exist, hidden in the literature. The seasonal succession of subtidal epipelton is to be fully documented.

Section 4.4.2 describes the distribution of epipelton at a range of depths in the sediments of Loch Goil. However, this study is limited in that they describe the algae only in the summer period. The study of the temporal pattern of epipellic algal succession provides an extra dimension to the understanding of the ecology of epipelton in Loch Goil and is fundamental to the principal aims of my research (aims: A, B and C; section 1.3).

The aim of this monitoring programme was to describe the seasonal variability of the epipelton within Loch Goil. The determination of the relationships and causal factors governing this seasonality is of secondary importance since only a very limited number of physical parameters (light attenuation, salinity and temperature) were monitored.

